CROWTH ARD AGAR CHARACTERISTICS OF SELECTED Gracilaria SPECIES UNDER DIFFERENT CULTIVATION METHODS

FORG CHUEN FAR

MASTER OF SCIENCE URIVERSITE MALAYSIA TERFNGGANU 2008

PDF processed with CutePDF evaluation edition www.CutePDF.com

1100068329

tesis



1100068329

Perpustakaan Sultanah Nur Zahirah (UMT) Universiti Malaysia Terengganu



Growth and agar characteristics of selected Gracilaria species under different cultivation methods / Fong Chuen Far.

	110006	8329
	_	
	· · ·	
•		

HAK MILIK PERPUSTAKAAN SULTANAH NUR ZAHIRAH IIMT

GROWTH AND AGAR CHARACTERISTICS OF SELECTED Gracilaria SPECIES UNDER DIFFERENT CULTIVATION METHODS

FONG CHUEN FAR

. . .

MASTER OF SCIENCE UNIVERSITI MALAYSIA TERENGGANU

2008

GROWTH AND AGAR CHARACTERISTICS OF SELECTED Gracilaria SPECIES UNDER DIFFERENT CULTIVATION METHODS

FONG CHUEN FAR

·. .

Thesis Submitted in Fulfilment of the Requirements for the Degree of Master of Science in the Faculty of Maritime Studies and Marine Science Universiti Malaysia Terengganu

Jan 2008

TABLE OF CONTENTS

Conte	ents			Page
APPR	OVA	L		iv
DECI	LARA	TION		vi
ACKNOWLEDGEMENT				
LIST	OF T	ABLES		ix
LIST	OF FI	GURES		xiii
LIST	OF A	BBREV	IATIONS	xviii
ABST	RAC	Т		XX
ABST	RAK			xxii
СНА	PTER			
c.m.		6.1.2		
1	INTF	RODUC	TION	1
2	LITE	DATID	RE REVIEW	6
2	2.1		aria Resources	6 6
	2.1	2.1.1		
			Distribution	6 7
			Reproduction and Life History	8
	2.2		tion of <i>Gracilaria</i>	10
	2.3		aria Cultivation and Production	10
	2.4		e methods of <i>Gracilaria</i>	13
	2.1		Field Culture	13
			Pond Culture	13
		2.4.3		16
	2.5	Agar		10
		2.5.1	Agar Quality	18
		2.5.2	Agar Properties	21
	2.6		ations of Agar	24
3	Grou	th and D	Productivity in Pond Culture	24
5	3.1		als and Methods	26 27
	5.1		Culture Species Description	
		3.1.2	Study Site Description	27 28
		3.1.3	Line Method	30
		3.1.4	Floating – cage Method	30
		3.1.5	Growth Measurement	33
		3.1.6	Productivity	34
		3.1.7	Harvest and Drying	35
		3.1.8	Water Parameters	36
		3.1.9	Statistical Analysis	37
	3.2	Results	-	38
	5.2	3.2.1	Pond Water Parameters	38
		3.2.2	Growth Rates	42

		3.2.3	Producti	vity	44	
		3.2.4	Percenta	ge of Water Content and Dry Weight	45	
		3.2.5		ge of Contaminants	47	
	3.3	Discus	ssions		48	
		3.3.1	Growtha	and Productivity	48	
		3.3.2	Post Har	vest	51	
		3.3.3	Fouling	organisms	52	
	3.4		nmendation		53	
	3.5	Conch	usion		54	
4	Gro	Growth and Productivity in Field Culture				
	4.1					
		4.1.1	Culture S	Species Description	57	
		4.1.2		te Description	58	
		4.1.3		oottom Rope and Netting Substrates	59	
		4.1.4		Measurement	63	
		4.1.5	Producti		64	
				and Drying	64	
			Water Pa		65	
		4.1.8	Statistica	al Analysis	65	
	4.2	Result			66	
		4.2.1	Field Wa	ater Parameters	66	
		4.2.2	Sporeling	g Densit y	69	
		4.2.3	-	Productivity	73	
				ge of Water Content and Dry Weight	75	
		4.2.5		ge of Contaminants	76	
	4.3	Discus		-	77	
		4.3.1	Sporeling	g Density and Productivity	77	
		4.3.2	Post Har	vest	79	
		4.3.3	Opportu	nistic Macroalgae and Invertebrates	79	
	4.4	Recon	mendation	ns	82	
	4.5	Conch	usion		83	
5	AGA	AGAR YIELD AND QUALITY ASSESSMENTS				
	5.1	Materi	als and Me	ethod	86	
		5.1.1	Alkali Tı	reatment and Agar Extraction	86	
		5.1.2	Determin	nation of Agar Quality	87	
			5.1.2.1	Determination of Gel Strength	87	
			5.1.2.2	Determination of Gelling Temperature	87	
			5.1.2.3	Determination of Melting Temperature	87	
			5.1.2.4	Fourier Transformed Infrared Spectroscopy (FTIR)	88	
		5.1.3	Statistica	l Analysis	88	
	5.2	Result			90	
		5.2.1	Agar Yie	eld	90	
		5.2.2	-		94	

		5.2.3	Gelling Temperature	97
		5.2.4	Melting Temperature	100
		5.2.5	Agar Content Assessments	103
	5.3	Discus	ssions	107
		5.3.1	Agar Yield	107
		5.3.2	Agar Quality	108
		5.3.3	Agar Content	110
	5.4	Recon	nmendations	112
	5.5	Conch	usion	113
5	GEN	IERAL	DISCUSSION	114
	GEN	IERAL	CONCLUSION	119
	REF	ERENC	CES	120
	APP	ENDIC	ES	130
	BIO	DATA (OF AUTHOR	163

I certify that an Examination Committee has met on 15th January 2008 to conduct the final examination of Fong Chuen Far on her Master of Science thesis entitled "Growth and Agar Characteristics of Selected *Gracilaria* Species Under Different Cultivation Methods" in accordance with the regulations approved by the Senate Unversity of Malaysia Terengganu. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follow:

Nor Antonina Abdullah, Ph.D. Faculty of Maritime Studies and Marine Science Universiti Malaysia Terengganu (Chairperson)

Siti Aishah Abdullah @ Christine A. Orosco, Ph.D. Faculty of Maritime Studies and Marine Science Universiti Malaysia Terengganu (Member)

Mohd. Azmi B. Ambak, Ph.D. Professor Faculty of Agrotechnology and Food Science Universiti Malaysia Terengganu (Member)

Mohamed Kamil B. Abdul Rashid, Ph.D. Associate Professor Faculty of Maritime Studies and Marine Science Universiti Malaysia Terengganu (Independent Examiner)

> MOHD. AZMTE AMBAK, Ph.D. Professor/Dean o Graduate School Universiti Malaysia Terengganu

Date: 12 June 2008

This thesis submitted to the Senate of Universiti Malaysia Terengganu and has been accepted as fulfillment of the requirement for the degree of Master of Science.

MOHD. AZMI B. AMBAK, Ph.D. Professor/Dean of Graduate School Universiti Malaysia Terengganu

. . .

Date: 12 June 2008

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UMT or other institutions.

FONG CHUEN FAR

Date: 12 June 2008

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to my supervisor, Dr. Siti Aishah Abdullah @ Christine A. Orosco for her trust and confidence in me, bringing me under her wings of guidance patiently. Prof. Dr. Mohd. Azmi B. Ambak for his understanding and review.

Mr. Subramaniam S/O Kathamuthu and Dr. Mohd. Fariduddin Othman of the Brackish Water Aquaculture Research Center for the cooperation and use of the culture pond. Special thanks to the staff at the Peladang, who had provided us shelter to conduct our works at the Research Center. Ms. Maslinda Kammaruddin for her assistance in the monthly monitoring works.

Dr. Ahemad Sade, Head of Division of Marine and Resources, Fisheries Research Centre Likas and Mr. Awang Hj. Pakar, Deputy District Director of Fisheries in Sandakan, for their support in the project in Sandakan; Mr. Ibnosion Abah and family, for their hospitality and assistance throughout my visits to Sandakan.

This research was funded by the Ministry of Science, Technology and Innovation (MOSTI) for the project on Genomic Approaches to Seaweed Genes and Natural Product Discovery (06-02-02-003 BTk/ER/016). Sub – project 2.2 Strain Selection and Cultivation of *Gracilaria* (Rhodophyta) in Malaysia.

Special thanks to Dr. Chuah Tse Seng for his helpful discussion and advise on the statistical analysis test.

Staff of the Netloft especially Mr. Fadzli, Mr. Mohd. Ibrahim and Mr. Adnan for their help in the set up of culture methods in Gelang Patah and Sandakan. Mr. Jamaluddin of the Chemistry laboratory and Ms. Nasrenim of Food Science laboratory for their guidance. Encouragement from the assistants of the Biodiversity laboratory is greatly appreciated.

Heartfelt thank to Mr. Chan Kian Weng who always there to help especially with the set up in Sandakan and Mr. Clenden Flant Langganggon for the three months of hard work in Gelang Patah. To all my friends who had shared my ups and downs throughout my study in Terengganu, I cherish every moment we spent together.

My utmost gratitude goes to my beloved parents, sister and brother. Your love, support and understanding helped me through my journey.

~Thank You~

LIST OF TABLES

No.	Title	Page
1	Quantity of agar extracted from <i>Gracilaria</i> in different culture methods.	20
2	Peaks found in phycocolloid infrared spectra with their attributed bonds (after Chopin <i>et al.</i> , 1999).	23
3	Comparison of nutrient measurements using standard colourmetric method with spectrophotometer and field kit (RQflex plus).	36
4	Comparison of the relative growth rate (RGR, % day ⁻¹) of <i>Gracilaria</i> species cultured in pond using line method in this study with similar study done by other researchers.	48
5	Mean biomass of G. changii on one fixed – bottom rope and netting substrate assessed in November 2005. The average biomass values are presented in wet and dry weight.	73
6	Mean productivity (in wet and dry weight) of <i>G. changii</i> on fixed – bottom rope and netting substrates assessed in November 2005.	73
A1.1	The cost of materials for each individual replica of method for pond culture.	130
A1.2	Water parameters and nutrient concentration in pond throughout the culture period.	131
A1.3	Two – way ANOVA test for comparison of growth between cultured species and methods in pond culture.	132
A1.4	RGR of <i>Gracilaria</i> sp. cultured in pond using line method from September 2004 to June 2005.	133
A1.5	Correlation tests between RGR of cultivars in both culture methods, water parameters and nutrient concentrations in pond culture.	135
A1.6	RGR of <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured in pond using floating – cage method from September 2004 to June 2005.	136
A1.7	Two – way ANOVA for comparison of percentage of water content in <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured in line and floating – cage methods.	137

Table

Pa	ge
----	----

A1.8	Two – way ANOVA for comparison of percentage of dry weight in <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured in line and floating – cage methods.	137
A1.9	Percentage of water content, dry weight and contaminant of $Gracilaria$ sp. and G . manilaensis in line and floating – cage methods throughout the culture period.	138
A1.10	Correlation tests between RGR of cultivars in both culture methods in pond and percentage of water content and dry weight.	139
A1.11	Two – way ANOVA for comparison of percentage of contaminants on <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured in line and floating – cage methods.	139
B1.1	The cost of materials per replica of substrate for field culture.	140
B1.2	Water parameters and nutrient concentrations in Sg. Kayu, Sandakan, Sabah throughout the culture period.	141
B1.3	Scheirer – Ray – Hare test for comparison of sporeling density between set up phases and culture substrates.	141
B1.4	Sporeling density (m ⁻²) on fixed – bottom rope substrates in field culture from September 2004 to June 2005.	142
B1.5	Correlation tests between sporeling density on both culture substrates, water parameters and nutrient concentrations in field culture.	144
C1.1	Scheirer – Ray – Hare test for comparison of agar yield of <i>Gracilaria</i> sp.and <i>G. manilaensis</i> in line and floating – cage methods extracted without and with alkali treatment at 90°C.	145
C1.2	Agar yield of <i>Gracilaria</i> sp., <i>G. manilaensis</i> and <i>G. changii</i> in pond and field culture. The agars were extracted with and without (native) alkali treatment.	146
C1.3	Correlation tests between agar yield of <i>Gracilaria</i> species in both culture methods, water parameters and nutrient concentrations in pond culture.	147

Table

C1.4	Two – way ANOVA for comparison of gel strength of agar from <i>Gracilaria</i> sp. and <i>G. manilaensis</i> in line and floating – cage methods extracted without and with alkali treatment at 90°C.	148
C1.5	Gel strength of <i>Gracilaria</i> sp., <i>G. manilaensis</i> and <i>G. changii</i> in pond and field culture extracted with and without (native) alkali treatment.	149
C1.6	Correlation tests between gel strength of <i>Gracilaria</i> species in both culture methods, water parameters and nutrient concentrations in pond culture.	150
C1.7	Two – way ANOVA for comparison of gelling temperature of agar from <i>Gracilaria</i> sp. and <i>G. manilaensis</i> in line and floating – cage methods extracted without and with alkali treatment at 90°C.	151
C1.8	Gelling temperature of agar from $Gracilaria$ sp. and G . manilaensis in line and floating – cage methods extracted with and without (native) alkali treatment.	152
C1.9	Correlation tests between gelling temperature of <i>Gracilaria</i> species in both culture methods, water parameters and nutrient concentrations in pond culture.	153
C1.10	Gelling and melting temperature of agar from G. changii harvested from P1 and P2 fixed – bottom rope and netting substrates in November 2005.	154
C1.11	Two – way ANOVA for comparison of melting temperature of agar from <i>Gracilaria</i> sp. and <i>G. manilaensis</i> in line and floating – cage methods extracted without and with alkali treatment at 90°C.	155
C1.12	Melting temperature of agar from $Gracilaria$ sp. and G . manilaensis in line and floating – cage methods extracted with and without (native) alkali treatment.	156
C1.13	Correlation tests between melting temperature of <i>Gracilaria</i> species in both culture methods, water parameters and nutrient concentrations in pond culture.	157
C1.14	Ratio absorbance of FT-IR spectra detected in agar from <i>Gracilaria</i> sp. and <i>G. manilaensis</i> in line method extracted with and without (native) alkali treatment.	158

Table

- C1.15 Ratio absorbance of FT-IR spectra detected in agar from Gracilaria 159 sp. and G. manilaensis in floating - cage method extracted with and without (native) alkali treatment.
- C1.16 Ratio absorbance of FT-IR spectra detected in agar from G. changii 160 in field culture extracted with and without (native) alkali treatment.
- C1.17 Scheirer Ray Hare test for comparison of ratio of absorbance at 161 890/2920 cm⁻¹ of agar from Gracilaria sp. and G. manilaensis in line and floating - cage methods extracted without and with alkali treatment at 90°C.
- C1.18 Scheirer Ray Hare test for comparison of ratio of absorbance at 161 930/2920 cm⁻¹ of agar from Gracilaria sp. and G. manilaensis in line and floating - cage methods extracted without and with alkali treatment at 90°C.
- C1.19 Scheirer - Ray - Hare test for comparison of ratio of absorbance at 162 1370/2920 cm⁻¹ of agar from Gracilaria sp. and G. manilaensis in line and floating - cage methods extracted without and with alkali treatment at 90°C.
- Scheirer Ray Hare test for comparison of ratio of absorbance at 1640/2920 cm⁻¹ of agar from *Gracilaria* sp. and *G. manilaensis* in C1.20 162 line and floating - cage methods extracted without and with alkali treatment at 90°C.

xii

LIST OF FIGURES

No.	Title	Page
1	Triphasic life cycle of Gracilaria (Critchley, 1993).	9
2	G. manilaensis (A) and Gracilaria sp. (B) cultured in line and floating – cage method in pond culture.	28
3	Map showing location of study site at Gelang Patah, Johore.	29
4	Illustration of line (A) and floating – cage (B) methods implemented in pond culture.	31
5	Line (A and B) and floating – cage (C and D) culture methods implemented in pond culture and distribution of the substrates in the pond in Brackish Water Aquaculture Research Center, Gelang Patah, Johore.	32
6	Mean light intensity (\pm SD) of the surface (Top) and bottom (Bottom) water of the culture pond. The light intensity was measured at monthly intervals from August 2004 to June 2005.	38
7	Water parameters measured in the culture pond at PPTAP, Johore. Plotted values represent the mean \pm SD at monthly intervals from August 2004 to June 2005.	40
8	Nutrient concentrations measured in the culture pond at PPTAP, Johore Plotted values represent the mean \pm SD at monthly intervals from August 2004 to June 2005.	41
9	Mean relative growth rate (% day ⁻¹ ; \pm SD) of <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured in pond using line method (A) and floating – cage (B) methods. The cultivars were measured at monthly intervals from September 2004 to June 2005.	43
10	Productivity of <i>Gracilaria</i> sp. and <i>G. manilaensis</i> cultured using line (A) and floating – cage (B) methods in pond from September 2004 to June 2005.	44
11	Percentage of water content and dry weight of <i>Gracilaria</i> sp. (A) and <i>G. manilaensis</i> (B) harvested monthly in line method from September 2004 to June 2005.	46
12	Percentage of water content and dry weight of <i>Gracilaria</i> sp. (A) and <i>G. manilaensis</i> (B) harvested monthly in floating – cage method from September 2004 to June 2005.	46

13	Percentage of contaminants on <i>Gracilaria</i> sp. and <i>G. manilaensis</i> harvested monthly in both line (A) and floating $-$ cage (B) methods from September 2004 to June 2005.	47
14	Gracilaria changii plant growing naturally in the intertidal area in Sungai Kayu, Sandakan, Sabah.	57
15	Map showing study area at Sungai Kayu, Sandakan, Sabah.	58
16	Fixed – bottom rope (A) and netting (B) substrates for spores settling in Sungai Kayu, Sandakan, Sabah.	60
17	Fixed – bottom rope (A) and netting (B) substrates for spore settling at the culture site. The culture substrates were distributed alternately in the intertidal areas in Sungai Kayu, Sandakan, Sabah (C and D).	61
18	Distribution setting of fixed – bottom rope (square with lines) and fixed – bottom netting (empty square) substrates for field culture trial in Sungai Kayu, Sandakan, Sabah. Phase 1 substrates were set up in June 2004 and Phase 2 substrates in August 2004.	62
19	Water parameters measured at Sungai Kayu, Sandakan, Sabah from June 2004 to April 2005. Plotted values represent the mean \pm SD at monthly intervals.	67
20	Nutrients measured at Sungai Kayu, Sandakan, Sabah measured from June 2004 to April 2005. Plotted values represent the mean \pm SD at bimonthly intervals.	68
21	Sporelings of G. changii attached on P2 fixed – bottom rope substrates (A and B) in October 2004. The size of the sporelings ranged from a discoid disc to approximately 1.5 cm in length.	70
22	Mean sporeling density (cm ⁻² \pm SD) of <i>G. changii</i> on fixed – bottom rope substrates measured at bimonthly intervals from August 2004 to June 2005 in Sungai Kayu, Sandakan, Sabah.	71
	Substrates in phase 1 (P1) were set up in June 2004 and phase 2 (P2) in August 2004. The growth curve (solid line) is fitted using polynomial regression.	
23	Mean sporeling density (cm ⁻² \pm SD) of <i>G. changii</i> on fixed – bottom netting substrates measured at bimonthly intervals from August 2004 to June 2005 in Sungai Kayu, Sandakan, Sabah. Substrates in phase 1 (P1) were set up in June 2004 and phase 2 (P2) in August 2004. The growth curve (solid line) is fitted using polynomial regression.	72

- Page
- Biomass production of G. changii on P1 (A) and P2 (B) fixed 74
 bottom rope substrates and P1 (C) and P2 (D) of fixed bottom
 netting substrates in Sungai Kayu, Sandakan, Sabah in November 2005.
- Percentage water content and dry weight of G. changii harvested
 from fixed bottom rope (blue) and netting (red) substrates in
 November 2005.
- Percentage of contaminants of G. changii harvested from fixed 76
 bottom rope (blue) and netting (red) substrates in November 2005.
- Opportunistic macroalga, *Padina* sp. (A), invertebrates including 81 colourful sponges (B, C and D), gastropod *Alvania sp.* (E) and barnacles *Balanus* sp. attached on fixed bottom ropes and netting substrates in field culture in Sungai Kayu, Sandakan, Sabah.
- 28 Base line method for determining transmittance (after Rochas et 89 al., 1986).
- Agar yield (%) from Gracilaria sp. (A) and G. manilaensis (B)
 cultured in line method in pond culture from September 2004 to
 June 2005. Agar was extracted without (●) and with alkali
 treatment of 5% NaOH at 90°C (■).
- Agar yield (%) from Gracilaria sp. (A) and G. manilaensis (B)
 cultured in floating cage method in pond culture from
 September 2004 to June 2005. Agar was extracted without (●)
 and with alkali treatment of 5% NaOH at 90°C (■).
- Agar yield (%) of G. changii harvested from P1 and P2 of fixed –
 bottom rope (A) and netting substrates (B) in field culture. Agar was extracted without (native) and with alkali treatment of 5% NaOH at 60°C, 70°C, 80°C and 90°C.
- Gel strength (g·cm⁻²) of agar extracted from Gracilaria sp. (A) 95 and G. manilaensis (B) cultured in line method in pond culture from September 2004 to June 2005. Agar was extracted without (●) and with alkali treatment of 5% NaOH at 90°C (■).
- Gel strength (g·cm⁻²) of agar extracted from Gracilaria sp. (A) 95 and G. manilaensis (B) cultured in line method in pond culture from September 2004 to June 2005. Agar was extracted without (●) and with alkali treatment of 5% NaOH at 90°C (■).

- Gel strength (g·cm⁻²) of G. changii harvested from P1 and P2 of fixed bottom rope (A) and netting substrates (B) in field culture. Agar was extracted without (native) and with alkali treatment of 5% NaOH at 60°C, 70°C, 80°C and 90°C.
- Gelling temperature (°C) of agar extracted from Gracilaria sp.
 (A) and G. manilaensis (B) cultured in line method in pond culture from September 2004 to June 2005. Agar was extracted without (●) and with alkali treatment of 5% NaOH at 90°C (■).
- Gelling temperature (°C) of agar extracted from Gracilaria sp.
 (A) and G. manilaensis (B) cultured in floating cage method in pond culture from September 2004 to June 2005. Agar was extracted without (●) and with alkali treatment of 5% NaOH at 90°C (■).
- Gelling temperature (°C) of G. changii harvested from P1 and P2 101 of fixed bottom rope (A) and netting substrates (B) in field culture. Agar was extracted without (native) and with alkali treatment of 5% NaOH at 60°C, 70°C, 80°C and 90°C.
- 38 Melting temperature (°C) of agar extracted from Gracilaria sp. 101 (A) and G. manilaensis (B) cultured in line method in pond culture from September 2004 to June 2005. Agar was extracted without (O) and with alkali treatment of 5% NaOH at 90°C (■).
- Melting temperature (°C) of agar extracted from Gracilaria sp. 102
 (A) and G. manilaensis (B) cultured in floating cage method in pond culture from September 2004 to June 2005. Agar was extracted without (●) and with alkali treatment of 5% NaOH at 90°C (■).
- 40 Melting temperature (°C) of *G. changii* harvested from P1 and P2 104 of fixed – bottom rope (A) and netting substrates (B) in field culture. Agar was extracted without (native) and with alkali treatment of 5% NaOH at 60°C, 70°C, 80°C and 90°C.
- 41 Ratio of absorbance of agar extracted without and with alkali 104 treatment at 90°C from *Gracilaria* sp. (A) and *G. manilaensis* (B) cultured in line method and the same species cultured in floating cage method (C and D, respectively) in pond from September 2004 to June 2005.
- Ratio of absorbance of agar of G. changii harvested from P1 and 105
 P2 fixed bottom rope substrates in field culture in November 2005. The agar was extracted without (native) and with alkali treatment at 60°C, 70°C, 80°C and 90°C.

96

98

- Ratio of absorbance of agar of G. changii harvested from PI and 104
 P2 fixed bottom netting substrates in field culture in November 2005. The agar was extracted without (native) and with alkali treatment at 60°C, 70°C, 80°C and 90°C.
- FTIR spectra of the commercially available agar powder from 106 Thailand, agar strip from China, agar from G. changii on P2 fixed
 bottom netting substrates without (Native) and with alkali treatment at 90°C (90°).

Page

LIST OF ABBREVIATIONS

Ø
mm
cm
m
in
cm ⁻¹
cm ⁻²
m ⁻¹
m ²
mm·s ⁻¹
wt
Qty
g
kg
t
g·cm ⁻²
g·m ⁻²
kg·cm ⁻³
°C
°C·min ⁻¹
min
h
RGR
% day ⁻¹
g dry wt ha ⁻¹
kg dry wt∙ha ⁻¹
kg wet wt ha ⁻¹
g dry wt·ha ⁻¹ ·yr ⁻¹
t dry wt
t dry wt∙ha ⁻¹
t wet wt ha ⁻¹
t dry wt·ha ⁻¹ ·yr ⁻¹

diameter
millimeter
centimeter
meter
inch
per centimeter
per square centimeter
per meter
per square meter
millimeter per second
weight
quantity
gram
kilogram
tonne
gram per square centimeter
gram per square meter
kilogram per cubic centimeter
degree centigrade
degree centigrade per minute
minute
hour
Relative growth rate
percentage per day
gram per dry weight per hectare
kilogram dry weight per hectare
kilogram wet weight per hectare
gram per dry weight per hectare per year
tonne dry weight
tonne dry weight per hectare
tonne wet weight per hectare
tonne dry weight per hectare per year

xviii

%	percentage
ppt	part per thousands
mg·l ⁻¹	milligram per liter
ml	milliliter
w/v	weight per volume
µM·m ^{-₂} ·s ^{−1}	micromolar per square meter per second
DO	Dissolved oxygen
NH4	Ammonium
NO ₂	Nitrite
NO ₃ -	Nitrate
PO4 ³⁻	Phosphate
SD	Standard Deviation
NaOH	Sodium Hydroxide
H ₂ SO ₄	Sulfuric Acid
FTIR	Fourier Transformed Infrared
N	large sample size
df	Degree of freedom
Р	Power of probability
F	Ratio of between and within group variance
R ²	Power of two Pearson's correlation
r	Pearson's correlation
r _s	Spearman's rank order correlation

Abstract of thesis presented to the Senate of Universiti Malaysia Terengganu in fulfillment of the requirements for the degree of Master of Science

GROWTH AND AGAR CHARACTERISTICS OF SELECTED Gracilaria SPECIES UNDER DIFFERENT CULTIVATION METHODS

FONG CHUEN FAR

Jan 2008

Chairperson	:	Siti Aishah Abdullah @ Christine A. Orosco, Ph.D.
Member	:	Professor Mohd. Azmi B. Amhak, Ph.D.
Faculty	:	Maritime Studies and Marine Science

Culture of the commercially important agarophyte, Gracilaria using two different approaches was done. Cultivation via vegetative propagation of Gracilaria sp. and G. manilaensis was conducted in pond in Gelang Patah, Johore, using line and floating cage methods while natural spore recruitment was implemented in the wild population of G. changii in Sandakan, Sabah on fixed - bottom rope and netting substrates with Phase 1 (P1) from June 2004 to June 2005 and Phase 2 (P2) from August 2004 to June 2005. In pond culture, line method supported better growth and productivity of Gracilaria sp. and G. manilaensis. Monthly monitoring of relative growth rate showed maximum growth of Gracilaria sp. in September 2004 $(2.28\pm0.63\% \text{ day}^{-1})$ and G. manilaensis in November 2004 $(2.88\pm1.80\% \text{ day}^{-1})$ with the total production of 72.5 kg dry wt ha⁻¹ and 51.2 kg dry wt ha⁻¹, respectively. In field culture, P2 (18.9 \pm 5.7 cm⁻²) and P1 (16.7 \pm 3.5 cm⁻²) fixed – bottom rope substrates showed highest sporeling density in October 2004. Density of sporelings was higher on P2 fixed - bottom netting substrates compared to P1, ranging from 1.1 ± 0.2 cm⁻² to 4.0 ± 1.6 cm⁻². PI fixed – bottom rope substrates produced a maximum crop of 9.15 t wet wt ha⁻¹ while higher biomass production was recorded in P2 fixed -

bottom netting substrates (6.63 t wet wt ha⁻¹). Epiphytism and fouling organisms were major problems in both cultures.

Yield, quality and content of agar extracted with and without alkali treatment from the monthly harvested cultivars in pond culture, and *G. changii* harvested in November 2005 were assessed and compared with commercially available agar strip from China and agar powder from Thailand. Yield of agar alkali treated at 90°C from all *Gracilaria* species ranged from 18.6% to 38.1%. Highest gel strength of agar from *G. manilaensis* (781 g·cm⁻²) and *G. changii* in PI fixed – bottom rope substrates (852 g·cm⁻²) were higher compared to commercial agar strip. Gel strength of agar from *G. manilaensis* in line method was correlated to light intensity. FTIR spectra of 890 cm⁻¹, 930 cm⁻¹, 1370 cm⁻¹ and 1640 cm⁻¹ were detected in all agars extracted from the cultured species. The study showed that line method in pond culture and fixed – bottom rope substrates in field culture had the potential to be applied in commercial production. In terms of species selection, *G. manilaensis* and *G. changii* are appropriate for food grade agar production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Malaysia Terengganu sebagai memenuhi keperluan untuk ijazah Master Sains

PERTUMBUHAN DAN CIRI – CIRI AGAR DARIPADA RUMPAI LAUT, Gracilaria SPESIS DALAM TEKNIK PENGKULTURAN YANG BERBEZA

FONG CHUEN FAR

Jan 2008

Pengerusi : Siti Aishah Abdullah @ Christine A. Orosco, Ph.D. Ahli : Profesor Mohd. Azmi B. Ambak, Ph.D. Fakulti : Pengajian Maritim dan Sains Marin

Pengkulturan Gracilaria, agarofit yang mempunyai kepentingan komersial, telah dijalankan dengan mengaplikasikan dua pendekatan yang berbeza. Pengkulturan melalui pembiakan vegetatif bagi Gracilaria sp. dan G. manilaensis telah dijalankan di kolam di Gelang Patah, Johor, dengan menggunakan kaedah tali and sangkar terapung manakala rekrutasi spora secara semulajadi telah diaplikasikan di populasi liar G. changii di Sandakan, Sabah dengan subtrat tali dan jaring kekal - dasar pada Fasa 1 (P1) dari Jun 2004 hingga Jun 2005 dan Fasa 2 dari bulan August 2004 hingga Jun 2005. Pengkulturan di kolam, kaedah tali menyokong pertumbuhan dan produktiviti Gracilaria sp. dan G. manilaensis yang lebih baik. Pemantauan bulanan menunjukkan pertumbuhan maximum Gracilaria sp. pada bulan September 2004 (2.28±0.63% hari⁻¹) dan G. manilaensis pada November 2004 (2.88±1.80% hari⁻¹) dengan produktiviti keseluruhan masing – masing sebanyak 72.5 kg berat kering ha⁻¹ dan 51.2 kg berat kering ha". Pengkulturan di lapangan, P1 (18.9±5.7 cm") dan P2 (16.7±3.5 cm⁻²) substrat tali kekal – dasar menunjukkan kepadatan spora tertinggi pada bulan Oktober 2004. Kepadatan spora adalah lebih tinggi di substrat jaring kekal - dasar P2 berbanding dengan P1, dengan julat antara 1.1±0.2 cm⁻² dan 4.0±1.6 cm⁻². Substrat tali kekal - dasar P1 mampu menghasilkan tuaian maximum sebanyak 9.15

ton berat basah·ha⁻¹ sementara produktiviti biomass yang lebih tinggi tercatat pada substrat jaring kekal – dasar P2 (6.63 ton berat basah·ha⁻¹). Epifit dan organisma penghalang merupakan masalah yang perlu diatasi dalam kedua – dua kaedah pengkulturan.

Penghasilan, kualiti dan kandungan agar yang diekstrak dengan dan tanpa rawatan alkali daripada tuaian bulanan dari kolam dan *G. changii* yang dituai pada November 2005 telah dinilai dan dibandingkan dengan agar keping dari China dan serbuk agar dari Thailand yang boleh didapati di pasaran. Penghasilan agar daripada semua *Gracilaria* spesis yang dirawat dengan alkali pada suhu 90°C berada dalam julat antara 18.6% dan 38.1%. Nilai tertinggi kekuatan gel agar daripada *G. manilaensis* (781 g·cm²) dan *G. changii* pada substrat tali kekal – dasar PI (852 g·cm⁻²) adalah lebih tinggi berbanding dengan nilai yang diperolehi daripada agar keping komersial. Kekuatan gel agar daripada *G. manilaensis* dari kaedah tali menunjukkan korelasi dengan intensiti cahaya. Spektra FTIR 890 cm⁻¹, 930 cm⁻¹, 1370 cm⁻¹ dan 1640 cm⁻¹ telah dikesan pada semua agar yang diekstrak daripada spesis tanaman. Kajian ini menunjukkan bahawa kaedah tali di pengkulturan kolam dan substrat tali kekal – dasar di pengkulturan lapangan berpotensi untuk diaplikasikan untuk penghasilan secara komersial. Dari segi pemilihan spesis, *G. manilaensis* dan *G. changii* sesuai untuk penghasilan agar gred makanan.